ABSTRACT

There is disclosed a tetrasaccharide, a hexasaccharide, on octasaccharide and a decasaccharide all capable of inhibiting complement activation and having less that 33% of undigested native heparin’s anticoagulation activity.
LOW MOLECULAR WEIGHT HEPARIN 
FRAGMENTS AS INHIBITORS OF COMPLEMENT 
ACTIVATION

This application is a continuation of U.S. patent appli-
cation Ser. No. 717,213, filed Mar. 28, 1985, now aban-
donned.

BACKGROUND OF THE INVENTION

1 Field of the Invention 
Resistance to infectious disease is the result of both innate and acquired immunity. Innate immunity is nonspecific and involves the body's defense mechanisms such as a the skin barrier, secre-
tions, and the cough reflex, whereas acquired immunity is very specific and often involves the production of a 
specific response (antibody) to a specific stimulus (anti-
gen) which may be a foreign substance or a natural 
substance which has gone awry.

The complement system serves as one part of the 
immune system and is also activated both by specific 
substances (antibody) and by the nonspecific substances 
(e.g. yeast wall and pneumococcal cell wall). The comple-
ment response involves a series of biochemical reac-
tions in which one protein acts on another, which in 
turn acts on another and so on. These proteins are desig-
nated by a alphabetical abbreviations and numbers com-
monly accepted in the field of immunology (e.g. Cl, B, 
P). Ultimately, this cascade may lead to the entrapment 
and/or cytolysis (destruction) of the offending sub-
stance. There are a number of other mechanisms by 
which the body defends itself, but for the purposes of 
brevity and simplicity, these have been omitted here.

As with any body function, the immune response 
does not always work perfectly or in the best interests 
of the individual. Allergies, for example, are the result 
of an overzealous immune response and are often quite 
harmful to the individual. Organ transplant rejection, 
on the other hand, is an example of an appropriate 
immune response which is detrimental to the individual.

Similarly, there are a whole variety of autoimmune 
diseases which result from the immune response attack-
ing the individual's own cells.

These immune and autoimmune disease represent a 
class of disease which are often difficult to treat effec-
tively. Though there are a number of drugs which exist 
to treat them, none are adequate to control or inhibit 
complement activation. The inhibition of complement 
may be very important in a variety of these diseases 
because as the complement system becomes active, 
there may be destruction and removal of cells and sub-
stances, which is detrimental to the individual. It is 
useful, therefore, to have substances which can be ad-
ministered to patients to prevent active complement 
activation.

Diseases in which such a substance may be therapeu-
tically useful include paroxysmal nocturnal hemoglo-
bininurea, rheumatoid arthritis, (in which the sub-
stance might be administered directly to a joint capsule 
to prevent complement activation), and hereditary angi-
oidema, (in which deficiency of a complement control 
protein leads to very active complement consumption).

Heparin, a polydispers sulfate polysaccharide,
used primarily as a blood anticoagulant during many 
medical and surgical procedures. Although this is hepa-
rin's primary activity, it also displays other secondary 
activities such as its ability to inhibit complement activ-
ity. However, heparin's potency as an anticoagulant has 
made its use as a complement activation inhibitor prob-
lematic. The use of anticoagulant active heparin for the 
treatment of immune disorders can result in hemor-
hagic complications.

Heparin's anticoagulant activity has been demon-
strated to be associated with the presence in its structure 
of a specific sequence for the binding of antithrombin 
III. Once bound to heparin, antithrombin III can then 
inhibit a number of blood coagulation factors, and thus 
prevent coagulation of blood. Several approaches have 
been used to separate heparin's anticoagulant (primary) 
activity from its complement cascade inhibition (sec-
ondary) activity. The removal of this antithrombin III 
binding sequence from heparin permits the exploitation 
of this drug for the treatment of immune related disor-
ders. Such disease states for which this drug would be 
useful include septic shock, rheumatoid arthritis, lupus 
and systemic lupus erythematosus.

Heparin's anticomplement activity is mediated by 
binding to a variety of complement proteins and 
thereby regulates both the classical and alternate amplifi-
cation pathways. Heparin inhibits a portion of the 
complement cascade by inhibiting generation of the cell 
bound amplification pathway C3 convertases, C3b,Bb, 
C3b,Bb,P and C3b,Bb,Nef. It also acts as a complement 
inhibitor by interfering with the binding site on C3b for 
B. Furthermore, it prevents the consumption of B by D 
in the presence of C3b again indicating a direct action 
on C3b.

2 Prior Art

Heparin has been used for the last half century as an 
anticoagulant. Commercial heparin, with an average 
molecular weight of 10,000-14,000, also possesses a 
multiplicity of other biological activities including an 
ability to regulate complement activation. The struc-
ture-activity relationship for heparin is well understood 
for anticoagulant activity, but it is poorly understood 
with effect on the complement system. Moreover, hepa-
rin's anticomplement activity is herefore been coupled 
with its anticoagulant activity which often is an 
undesirable side effect.

Therefore, it is desirable to provide isolated compo-
nents of heparin which have high anticomplement ac-
tivity with low anticoagulant activity.

SUMMARY OF THE INVENTION

Depolymerized heparin is fractionated to recover a 
mixture of polysaccharide fragments. Further fraction-
ation on the basis of size permits the isolation of a tetra-
saccharide mixture, a hexasaccharide mixture, an octa-
saccharide mixture, a deca saccharide mixture and a 
mixture comprised of higher oligosaccharides. The 
tetrasaccharide mixture can be further fractionated into 
over 12 individual tetrascarachide components. The 
mixture of polysaccharide fragments and the size sepa-
rated fractions are all capable of inhibiting complement 
activation while all show less than 33% of undigestd 
native heparin's anticoagulant activity. An individual 
hexasulfated tetrascarachide having no detectable anti-
coagulant activity is disclosed which exhibits significant 
ability to inhibit complement activation. Said polysac-
charide fragments may be employed therapeutically to 
treat immune disorders involving activation of comple-
ment including autoimmune diseases.
BRIEF DESCRIPTION OF THE DRAWINGS

The accompanying figures and drawings are provided to demonstrate the results obtained in the specific examples illustrating the invention.

FIG. 1 represents fractionation of mixed depolymerized heparin fragments utilizing low pressure gel permeation chromatography.

FIG. 2a represents the results of further separation utilizing high performance liquid chromatography of the heparin derived mixtures of tetrascarrihides.

FIG. 2b represents the results of further separation utilizing high performance liquid chromatography of the heparin derived mixtures of hexascarrihides.

FIG. 2c represents the results of further separation utilizing high performance liquid chromatography of the heparin derived mixtures of octascarrihides.

FIG. 2d represents the results of further separation utilizing high performance liquid chromatography of the heparin derived mixtures of decascarrihides.

Note: The flow rate in FIGS. 2a, b, c and d is 1.5 ml/min, chart speed is 15 cm/hr, 0.2 a.u.f.s., with a linear sodium chloride gradient at pH of 3.5.

FIG. 3a represents the results of an assay of the tetrascarrihides mixture eluted in FIG. 2a plotting the component number against the percent of native heparin's anticomplement activity.

FIG. 3b represents the results of an assay of the hexascarrihides mixture eluted in FIG. 2b plotting the component number against the percent of native heparin's anticomplement activity.

FIG. 3c represents the results of an assay of the octascarrihides mixture eluted in FIG. 2c plotting the component number against the percent of native heparin's anticomplement activity.

FIG. 3d represents the results of an assay of the decascarrihides mixture eluted in FIG. 2d plotting the component number against the percent of native heparin's anticomplement activity.

FIG. 4 is a representative of the structural for the hexasulfated tetrascarrihide characterized in Example III.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The compositions of this invention are characterized by the capacity to inhibit complement activation, while having little or no anticoagulant activity. The compositions of this invention are obtained by depolymerizing native heparin with heparinase. The mixture of polysaccharides obtained by the heparinase digestion have the desired anticomplement activity and have an average molecular weight of between 900 and 1200 and range from 500 to 5000.

Other methods of heparin depolymerization may be utilized including chemical hydrolysis, chemical elimination cleavage, deaminative cleavage and oxidative cleavage.

Other compositions of this invention comprise fragments isolated from the above polysaccharide mixture. These fragments comprise:

- a mixture of tetrascarrihides having an average molecular weight of 1,180 and a chain size of 4;
- a mixture of hexascarrihides having an average molecular weight of 1,400 and a chain size of 6;
- a mixture of octascarrihides having an average molecular weight of 2,300 and a chain size of 8;
- a mixture of decascarrihides having an average molecular weight of 2,700 and a chain size of 10;
- a mixture of higher oligosaccharides having an average molecular weight of 4,000 and a chain length of >10.

After heparin has been depolymerized using either a free or immobilized heparinase, the enzyme is removed by filtration, precipitation, chromatography or some other standard technique known to the art. The recovered polysaccharide mixture is then concentrated by freeze-drying or precipitation. The fragments thus obtained are redisolved in a suitable aqueous solution such as IM ammonium bicarbonate and applied to a gel column at a high concentration, usually at 100 mg/ml. The column is packed with conventional chromatography gel beads of controlled pore size, such as Fractogel TSK 40 or 50, and then are eluted with a suitable aqueous solution such as 1М ammonium bicarbonate.

The column Effects separation of the fragments on the basis of size. Appropriate heparin and chondroitin derived carbohydrates of various well defined molecular weights are used as molecular weight standards. Further fractionation of the sized fragment mixtures is accomplished on the basis of charge using an anion exchanger, such as by high performance liquid chromatography on a column packed with strong anion exchanger. Such fractionation of the tetrascarrihides mixture resulted in the isolation of a hexasulfated tetrascarrihide (FIG. 4) having the appropriate anticomplement activity.

Similar fractionation has been accomplished on the other sized mixtures of fragments and in each case, individual components have been isolated with the appropriate anticomplement activity. In all cases these individual components, with high activity to inhibit complement activation, have a degree of sulfation higher than the average for the polysaccharide mixture as a whole.

The sized mixtures of fragments are also fractionated using affinity chromatography. Columns packed with Antithrombin III-Sepharose are used to remove residual traces of anticoagulant activity from the sized fragment mixtures. Alternatively, complement proteins coupled to Sepharose are used to enrich the sized fragment mixtures in their ability to inhibit complement activation.

The sized fragment mixtures, as sodium salts, are characterized with respect to molecular weight by gel chromatography against standards of known molecular weight. Additionally their average molecular weight is estimated from their absorbance at a fixed concentration using the known molar absorptivity for their chromophore. The structure of the hexasulfated tetrascarrihide (FIG. 4) was established by both proton and carbon nuclear magnetic resonance spectroscopy.

The degree of sulfation of individual components is established electrophoretically or chemically using a rhodizonate sulfate assay. The anticoagulant activity or absence thereof is determined using activated partial thromboplastin time and Factor Xa clotting assays as well as Thrombin-Antithrombin III and Factor Xa-Antithrombin III amidolytic assays. The capacity to inhibit complement activation is measured by the generation of the alternative pathway convertase, EAC4b,3b,Bb,F. The results of this activity assay is given in TABLE I and FIGS. 3a, b, c and d illustrated.

As noted above, the compounds of this invention can be isolated in the form of their sodium or ammonium
salt. However, it is to be understood that the compounds of this invention can be utilized in the form of the free acids, other metallic or non-metallic salts.

Representative suitable salts include alkaline earth metal bases, ammonium and alkyl substituted ammonium salts or the like. These salts can be formed by any conventional means.

**TABLE 1**

<table>
<thead>
<tr>
<th>SAMPLE</th>
<th>CONCENTRATION (µg/ml) GIVING 50% INHIBITION OF EACb, 3h, 8h, P CONVERTASE</th>
</tr>
</thead>
<tbody>
<tr>
<td>HEPARIN*</td>
<td>7</td>
</tr>
<tr>
<td>POLYSACCHARIDE</td>
<td>75</td>
</tr>
<tr>
<td>MIXTURE</td>
<td>85</td>
</tr>
<tr>
<td>TETRASACCHARIDE</td>
<td>75</td>
</tr>
<tr>
<td>HEXASACCHARIDE</td>
<td>75</td>
</tr>
<tr>
<td>MIXTURE</td>
<td>36</td>
</tr>
<tr>
<td>OCTASACCHARIDE</td>
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</tr>
<tr>
<td>MIXTURE</td>
<td>15</td>
</tr>
<tr>
<td>DECASACCHARIDE</td>
<td>15</td>
</tr>
<tr>
<td>LARGE OLIGOSACCHARIDE MIXTURE</td>
<td>5</td>
</tr>
<tr>
<td>HEXASULFATED TETRASACCHARIDE</td>
<td>50</td>
</tr>
</tbody>
</table>

*Heparin anticoagulant activity = 150 U/mg while the hexasulfated tetrasaccharide was <1 U/mg by all assays.

The following examples illustrate the present invention and are not meant to limit the same.

**EXAMPLE I**

Enzymatic Depolymerization of Heparin

Heparin fragments were prepared in a mixture of porcine mucosal heparin (50 mg/ml) and purified heparinase (heparin lyase EC 4.2.2.7) derived from Flavobacterium heparinum (5U/ml, 1 U = 1 µmol bonds cleaved/min), both in buffer consisting of 0.25 M sodium acetate and 0.0025 M calcium acetate at pH 7. This mixture was shaken at 30°C until the reaction reached completion. The pH of the mixture was then adjusted to pH 4 and the enzyme was removed by passage over a column containing SP-Sephadex C-50. The eluent containing the polysaccharide mixture was adjusted to pH 7 and freeze dried.

**EXAMPLE II**

Fractionation of Polysaccharide Compounds

The polysaccharide mixture obtained, as described in EXAMPLE I, was dissolved in water (500 mg in 2 ml) and was loaded onto a column packed with Fractogel TSK F40 (80 x 2.5 cm) and eluted with 1 M ammonium bicarbonate buffer at 0.3 ml/min. The eluent was diluted with water and its absorbance was measured at 232 nm (FIG. 1). Six distinct peaks were observed representing, from last to first eluted, disaccharide, mixed tetrasaccharide fragments, mixed hexasaccharide fragments, mixed octasaccharide fragments, mixed decasaccharide fragments and mixed higher oligosaccharide fragments. These peaks were identified on the basis of elution volume as compared to heparin and chondroitin derived standards of known molecular weight and by their absorbance at 232 nm based on a reported molar absorptivity of 5.2 x 10^5 M^-1 cm^-1.

Each sized fragment mixture (1 mg in 0.01 ml) was then applied to a strong anion exchange high performance liquid chromatography column and eluted using 180 ml linear gradient (Y + 1.5 x 10^-4 M/sec x + 0.2 M, sodium chloride at pH 3.5) and detected by absorbance at 232 nm (FIG. 2a-d). The resulting individual components were collected, dialyzed against 1000 volumes of water using controlled pore dialysis membranes (MW cutoff = 1,000) and freeze dried.

Further fractionation of either the polysaccharide sized mixture or the individual components separated by affinity chromatography involved the use of either immobilized antithrombin III or an immobilized complement protein such as C3b. These proteins were immobilized onto cyrogen bromide activated Sepharose by standard methods. The immobilized protein was then packed into a column equilibrated with 50 mM phosphate buffer (pH 7) and the fragment sample in the same buffer was applied. When using antithrombin III as the affinity matrix, the eluent was collected, dialyzed (as above), freeze dried and was then measured for biological activity. When using immobilized complement proteins, such as C3b as the affinity matrix, the eluent initially obtained in 50 mM buffer was discarded and the column was washed first with 5 column volumes of 50 mM buffer and then with 1 column volume of 2 M phosphate buffer (pH 7) to elute the fragments which initially bound to the column. These fragments were then dialyzed (as above), freeze dried and measured for biological activity.

**EXAMPLE III**

Structural Elucidation

The structure of the hexasulfated tetrasaccharide (FIG. 4), as well as the other individual fragment components was determined by using chemical, electrophoretic and spectroscopic methods. The degree of sulfation was determined using either: (1) the rhodizionate assay for inorganic sulfate after pyrolysis of the sample, or (2) by paper electrophoresis of the unmodified fragment. The number of uronic acid residues was measured using carbazole dye reagent. The precise structure of the hexasulfated tetrasaccharide was established using (90.6 MHz) 13C and (360 MHz) 1H nuclear magnetic resonance spectroscopy at 0.05 M in D2O.

**EXAMPLE IV**

Biological Assay

The ability of a fragment or mixture of fragments, at concentrations from 1-50 µg, to inhibit complement activation was determined by hemolytic assay. Half-isotonic veronal-buffered saline, pH 7.5 containing 0.1% gelatin (GVB), GVB containing 2.5% dextrose, 0.5 M magnesium and 0.15 mM calcium (DGVB+) and GVB containing 400 mM EDTA were used as buffers in the hemolytic assays.

Complement proteins C3 (Tack and Prahl, Biochemistry 15:4513), B (Hunsicker et al., J. Immunol 110:128), D (Fearon and Austen, J. Exp. Med. 142:856) and P (Fearon and Austen, Proc. Natl. Acad. Sci. USA. 74:1683) were purified to homogeneity and quantitated as described. C3b was generated from purified C3 as described previously (Gitlin et al., J. Exp. Med. 141:1221; Weiler et al., J. Exp. Med. 147:509). Rat serum, as a source of terminal components, was obtained from Rockland Inc. (Gilbertsville, PA.).

EACb, 3b cellular intermediates were prepared as described previously (Fearon et al., J. Exp. Med. 138, 1305; Lachmann and Hobart, Handbook of Experimental Immunology, 3rd Edn. Chapter 5a). Heparin and its fragments were examined for ability to inhibit generation of the amplification pathway conversase as previ-
We claim:

1. A compound derived from depolymerized heparin comprising a free acid or metallic or non-metallic salt of a tetrasccharide, a hexasccharide, an octasccharide, or a mixture thereof, wherein said compound has biological activity to inhibit complement activation, wherein said compound has diminished anticoagulant activity as compared to native heparin, and wherein the molecular weight of said compound is no greater than about 2300 daltons.

2. The compound of claim 1 wherein said compound is obtained by heparinase EC-4.2.2.7 digestion.

3. The compound of claim 1 wherein the compound is a tetrasccharide.

4. The compound of claim 3 wherein the tetrasccharide is a hexasulfated tetrasccharide.

5. The compound of claim 1 wherein the compound is a hexasccharide.

6. The compound of claim 1 wherein the compound is an octasccharide.

7. A compound derived from depolymerized heparin, comprising a free acid or metallic or non-metallic salt of a decescharide, said compound has biological activity to inhibit complement activation and has diminished anticoagulant activity as compared to native heparin.

8. A method of treating immune disorders in mammals, comprising the intravenous administration of the decescharide of claim 7.

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